

Lecture Program - per 30th May 2006 (subject to change)
**4th European Short Course on Principles & Applications of
Time-Resolved Fluorescence Spectroscopy,
Berlin, October 30-November 3, 2006**

Monday	Joseph R. Lakowicz: „Basic definitions and principles of fluorescence (1 and 2)“ (2 h 45 min, 9:30-10:45, and 11:15-12:45)
	<ul style="list-style-type: none"> ■ Jablonski diagram and stokes shift ■ Basic spectral properties ■ Excitation and emission spectra ■ Fluorescence anisotropy ■ Fluorescence lifetime ■ Energy transfer
	Rainer Erdmann: „Instrumentation (1)“ (1 h 30 min, 13:45–15:15)
	<ul style="list-style-type: none"> ■ Overview of steady state fluorometer design ■ Light sources: lamps, LEDs, lasers ■ Wavelength selection: monochromators, filters ■ Detectors: PMTs, MCP-PMT, SPAD, CCD ■ Electronics ■ Analog and photon counting ■ Design rules ■ Sources of error in fluorescence ■ Introduction to time domain measurement ■ Introduction to frequency domain measurement ■ Special considerations for NIR applications
	Matthias Patting : „Introduction to data analysis“ (30 min, 15:45-16:15)
	<ul style="list-style-type: none"> ■ Typical approaches in Time-Correlated Single Photon Counting (TCSPC) data analysis ■ Common artefacts and how to handle them ■ Spoiled data and how to avoid them ■ Choosing appropriate models ■ Step by step example
Tuesday	Zygmunt (Karol) Gryczynski / Andreas Bültner: „Introduction to Hands-on experiments“ (30 min, 16:15-16:45)
	<ul style="list-style-type: none"> ■ Physics behind the experiments
	Companies: „Introduction to Hands-on experiments“ (15 min per company, 16:45-18:15)
	<ul style="list-style-type: none"> ■ Instrumental aspects of the experiments
	Joseph R. Lakowicz: „Time-resolved fluorescence“ (1 h 45 min, 8:30-10:15)
	<ul style="list-style-type: none"> ■ Resolution of complex decays ■ Multi-exponential anisotropy decays ■ Transient effects in quenching ■ Time-Resolved Emission Spectra (TRES)
Tuesday	Michael Wahl: „Instrumentation (2) for time-correlated photon counting and fluorescence lifetime imaging“ (1 h 30 min, 10:45-12:15)
	<ul style="list-style-type: none"> ■ Advantages and difficulties of the TCSPC method ■ Modern excitation sources ■ Specifics of sample compartments and detection optics ■ Detectors for TCSPC ■ Compact photon counting electronics incl. multi-photon counting ■ Electronics for multidimensional TCSPC (including routers) ■ Electronics for Time-Tagged Time Resolved (T²R) data acquisition ■ TCSPC instrumentation for Fluorescence Lifetime Imaging (FLIM)
	Zygmunt (Karol) Gryczynski: „Analytical applications of fluorescence“ (1 h 45 min, 13:30-15:15)
	<ul style="list-style-type: none"> ■ Analytical determinations by fluorescence ■ Ratiometric determination based sensing ■ Anisotropy-based sensing ■ Fluorescence lifetime-based sensing ■ Modulation based sensing ■ Energy transfer-based lifetime sensing of metal ions ■ Visual polarization sensing ■ Error sources in fluorescence assays

Wednesday	<p>Joseph R. Lakowicz: „Time dependent phenomena“ (1 h 30 min, 8:30-10:00)</p> <ul style="list-style-type: none"> ■ Multi-exponential decays ■ Time domain lifetime measurements ■ Frequency domain lifetime measurements ■ Quenching: static, dynamic, transients ■ Anisotropy decays ■ Energy transfer – distance distribution ■ Time-dependent spectral relaxation ■ Excited state reactions
	<p>Johan Hofkens: „Modern fluorescence microscopy“ (1 h 30 min, 10:30-12:00)</p> <ol style="list-style-type: none"> 1. Hardware aspects: <ul style="list-style-type: none"> ■ Introduction to microscopy ■ Basics of (fluorescence)-microscopy (lightpath inside a microscope, light collection (lenses, objectives, aberration), resolution, focus properties, working distances, transmission / epi-illumination, excitation sources / possible detectors / general setup) ■ Widefield microscopy ■ Total Internal Reflection Fluorescence (TIRF) ■ Confocal and Multi-photon microscopy ■ High resolution microscopy (Stimulated Emission Depletion Spectroscopy (STED), 4-PI) ■ Spinning disk microscopy ■ Laser Scanning microscopy 2. Applications: <ul style="list-style-type: none"> ■ General imaging ■ Spectral mixing / unmixing ■ FLIM (phase, TCSPC, gating, ...) ■ Förster Resonance Energy Transfer (FRET) / Imaging FRET ■ Fluorescence Correlation Spectroscopy (FCS) ■ Deconvolution microscopy
Thursday	<p>Axel Dürkop: „Fluorescent markers, probes and labels“ (1 h 30 min. 8:30-10:00)</p> <ol style="list-style-type: none"> 1. Intrinsic fluorophores <ul style="list-style-type: none"> ■ Aromatic amino acids (Tyr, Trp, Phe) ■ Enzyme co-factors (NADH, FAD,...) ■ Tissues 2. Extrinsic fluorophores <ul style="list-style-type: none"> ■ Protein probes (Fluorescein, Rhodamines, Dansyl, BodiPy,...) ■ Membrane probes (DPH, TMA-DPH, Parinaric acid, Fatty acids,...) ■ DNA probes (Ethidium bromide, DAPI, Acridine Orange, Adenine, Guanine,...) 3. Chemical sensing probes <ul style="list-style-type: none"> ■ Ion indicators (MQAE, FURA, Calcium Green,...) ■ Fluorogenics probes (Fluorescamine, NBD-Cl,...) 4. Fluorescent Proteins <ul style="list-style-type: none"> ■ Phycobiliproteins ■ GFP, CFP, YFP,... 5. Lanthanides 6. Metal-Ligand complexes 7. Protein sensors 8. Luminescent nanoparticles aka Quantum Dots
	<p>Jörg Enderlein: „Fluorescence fluctuation and single molecule spectroscopy“ (1 h 30 min, 10:30-12:00)</p> <ol style="list-style-type: none"> 1. Physical principles of single molecule fluorescence spectroscopy <ul style="list-style-type: none"> ■ General properties of molecular light absorption and emission ■ Fluorescence lifetime and polarization ■ Single-pair Förster Resonance Energy Transfer (spFRET) 2. Fluorescence fluctuation spectroscopy <ul style="list-style-type: none"> ■ Confocal epi-fluorescence microscopy ■ Time-Tagged Time-Resolved photon counting ■ Fluorescence Correlation Spectroscopy (FCS) ■ Fluorescence Intensity Distribution Analysis (FIDA) ■ Single molecule burst analysis 3. Single Molecule Imaging <ul style="list-style-type: none"> ■ Wide-field fluorescence imaging microscopy ■ Single molecule tracking ■ Imaging single molecule orientations ■ Monitoring the interaction between individual molecules ■ Stoichiometry of molecular complexes

Friday	<p>Martin Hof: "Solvent relaxation techniques: Application in studies of biomolecules" (1 h 30 min, 13:00-14:30)</p> <ul style="list-style-type: none"> ■ Solvent relaxation (SR) and steady state spectra ■ Time-resolved emission spectra ■ SR in biomembrane research ■ Protein-Membrane interactions studied by SR ■ Lipid systems for drug delivery protocols studied by SR ■ SR in protein and DNA research
	<p>Matthias Patting: „Advanced data analysis“ (1 h 30 min, 8:30-10:00)</p> <ul style="list-style-type: none"> ■ Fundamentals of TCSPC fitting ■ Decay models ■ Advanced error analysis ■ Fluorescence Lifetime Imaging (FLIM) analysis ■ Förster Resonance Energy Transfer (FRET) analysis
	<p>Manfred Auer: „High throughput screening“ (2 h 15 min, 10:00-10:45 and 11:15-12:45)</p> <ul style="list-style-type: none"> ■ The drug discovery process ■ General aspects of high throughput screening ■ Ensemble averaging fluorescence technologies in high throughput screening ■ Single molecule spectroscopy technologies in high throughput screening ■ Affinity selection, chemical genomics, chemical genetics in drug discovery