

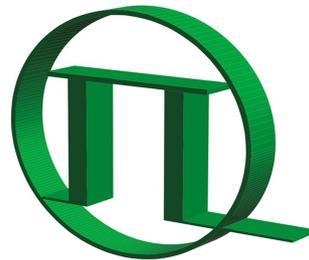
UV MicroTime 200

Crossing the Border towards Deep UV – Time-resolved Microscopy of Native Fluorophores

Marcelle König¹, Sebastian Tannert¹, Sandra Orthaus¹, Volker Buschmann¹,
Thomas Schönau¹, Kristian Lauritsen¹, Rainer Erdmann¹, and
Reinhild Beyreiss², Stefan Nagl², Detlev Belder²

¹ PicoQuant GmbH, Rudower Chausse 29, 12489 Berlin, Germany, www.picoquant.com

² University Leipzig, Institute of Analytical Chemistry, Linnéstr. 3, 04103 Leipzig, Germany



PICOQUANT

UV MicroTime 200, Webtalk 2013

Copyright of this document belongs to PicoQuant GmbH.

No parts of it may be reproduced, translated or transferred to third parties without written permission of PicoQuant GmbH.

© PicoQuant GmbH, 2013

Please note...

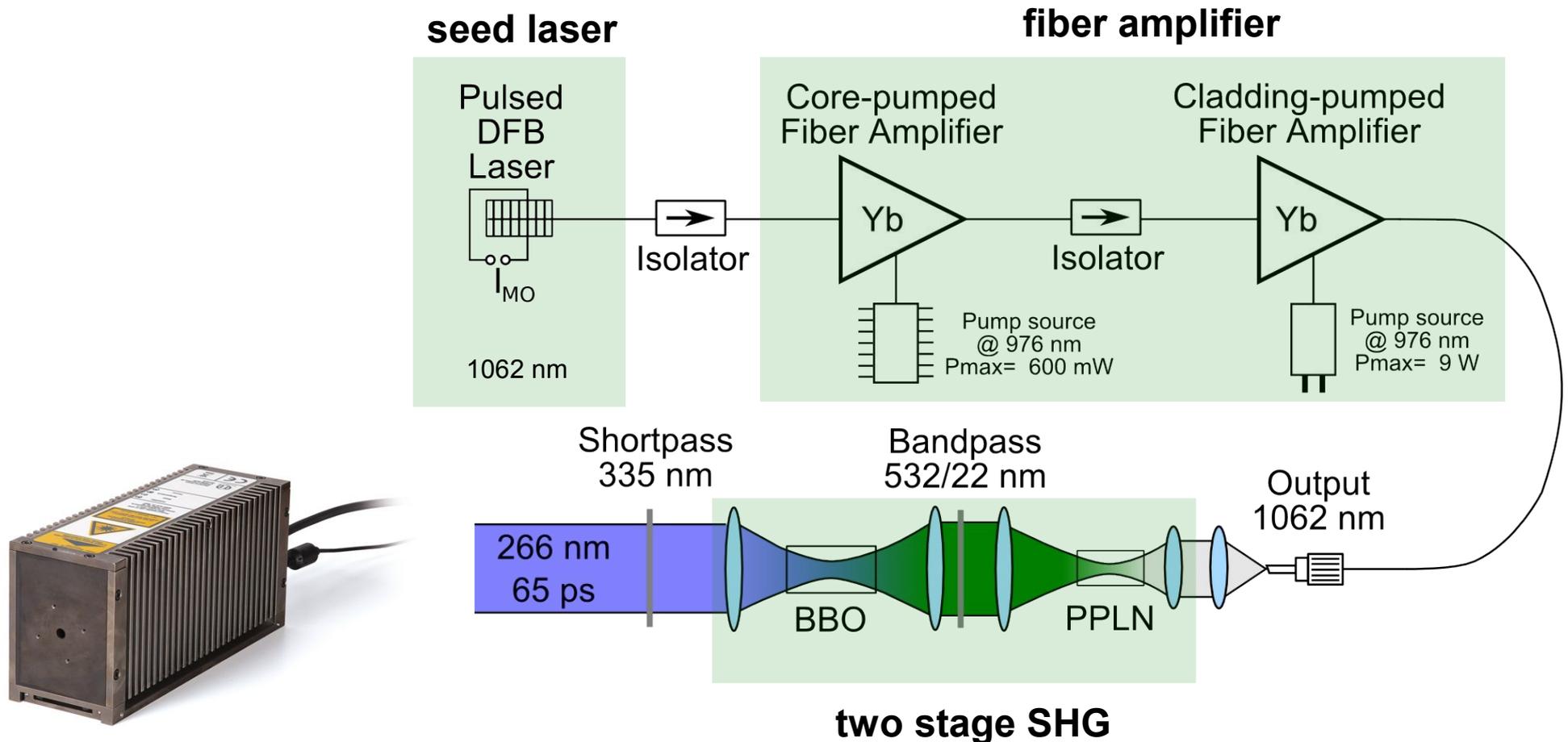
We at PicoQuant always try to present our latest findings to the scientific community as quickly as possible.

The creation of suited presentation material with in-depth information is, however, not always possible in a short time frame.

We have therefore decided that for those cases, it would be beneficial to the scientific community to make our presentations or parts of presentations, that where given on conferences, available to the public. As a consequence, please understand that it might be possible that sometimes information is missing to understand all information included in a slide.

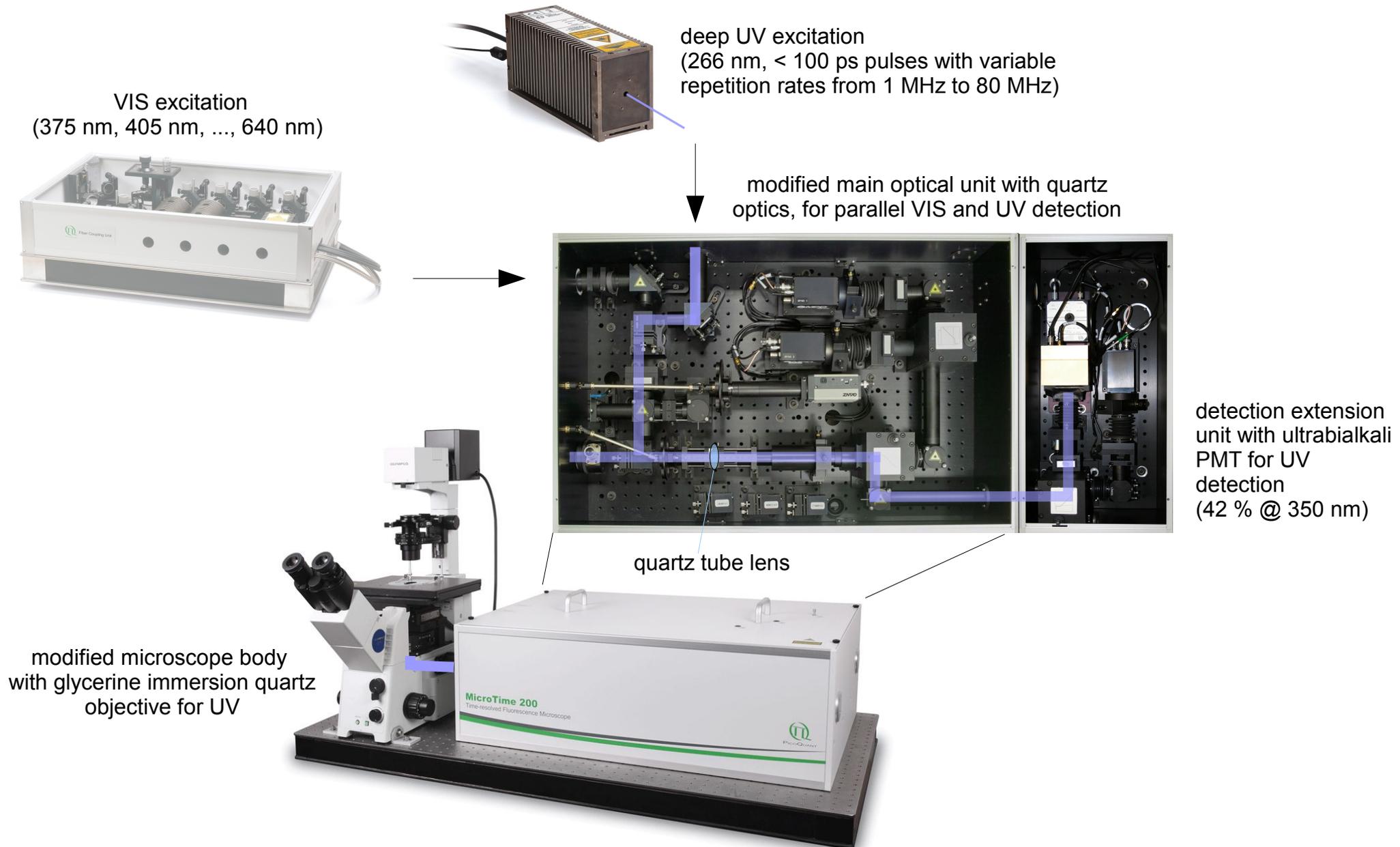
Don't hesitate to contact us in case you have any questions or need more information.

Fiber Amplified UV Laser for Pulsed Deep UV 266 nm Excitation



- 266 nm, < 100 ps pulses
- variable repetition rates from 1 MHz to 80 MHz

UV Configuration of the MicroTime 200



UV Microscopy Applications: Intrinsic Fluorescence

Label-free detection of molecules

Measuring intrinsic fluorescence from proteins

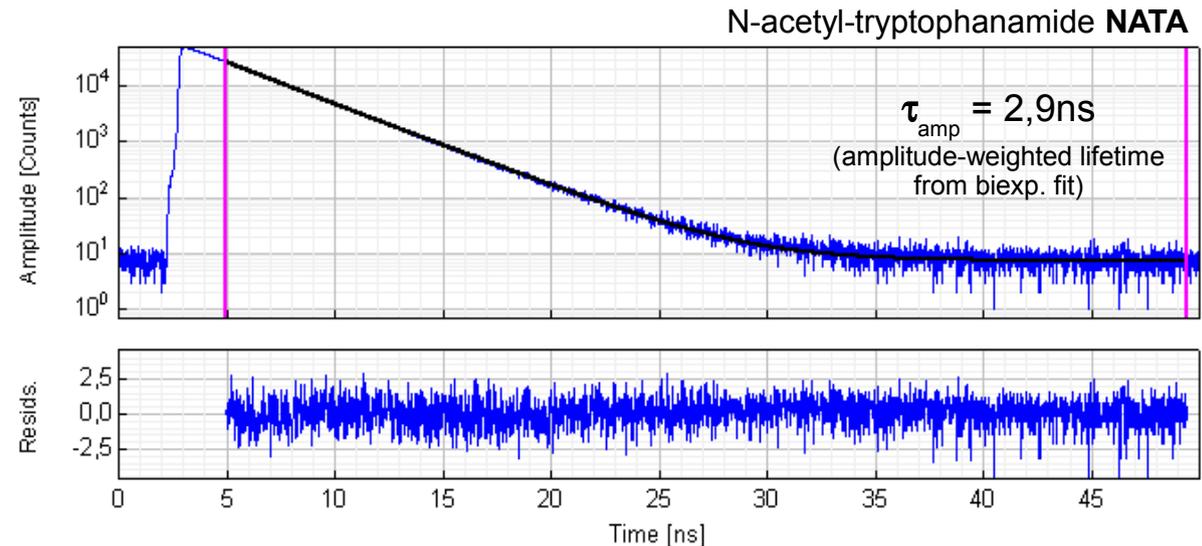
- native occurrence of detected species
- no labeling strategy needed
- low quantum efficiencies

Natural fluorophores

- excitation with 266 nm
→ aromatic amino acids in proteins:
 - tryptophan (e.g. NATA)
 - tyrosine
 - (phenylalanine)
- excitation with 355 nm
 - enzyme cofactors NADH/NADPH
 - collagen, elastin

Excitation of several natural fluorophores in biological samples

- identification with the aid of fluorescence lifetimes

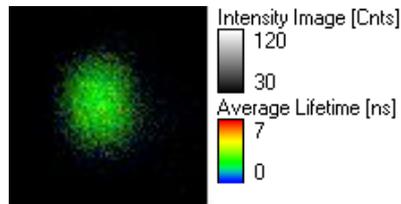


- Excitation: 266 nm (PicoQuant), 20 MHz
- Quartz objective 40x, NA 1.25, glycerin
- Optical filters: Z266RDC, longpass 300 nm
- Detection: ultrabialkali PMT

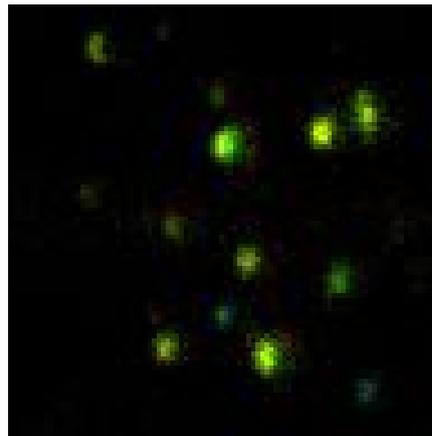
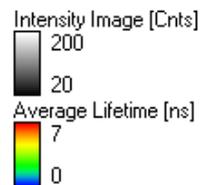
UV MicroTime 200: Label-free FLIM

266 nm grants access to the intrinsic fluorescence of tryptophan-containing proteins

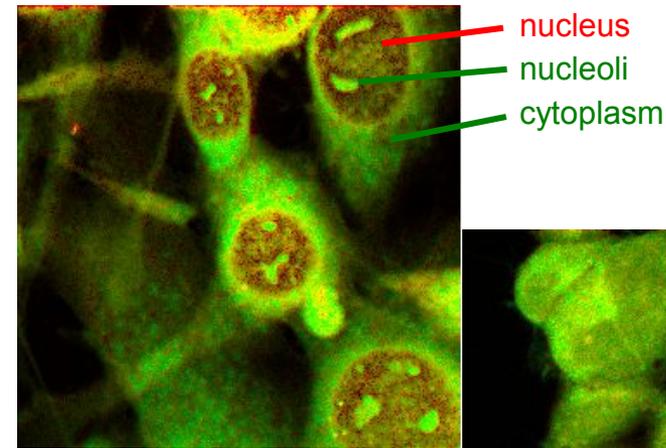
FLIM with streptavidin-coated beads
(\varnothing 500 nm, 1 streptavidin = 24 tryptophans),
immobilized on quartz coverslips



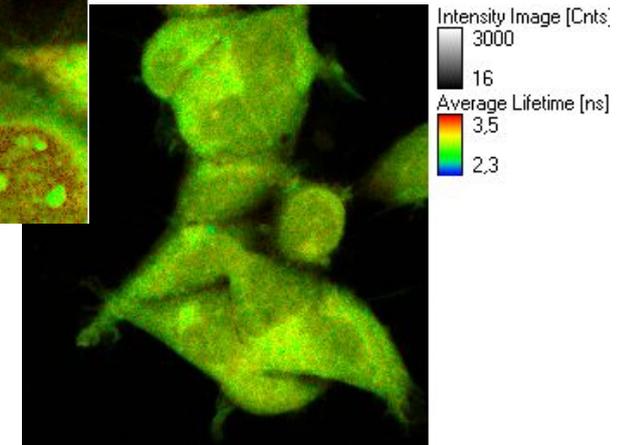
1.5 x 1.5 μ m



Label-free FLIM with biological cells
→ aromatic amino acids (mainly tryptophans)
within the proteins become visible



3T3-L1 cells, fixed
(methanol)
80 x 80 μ m



HEK293 cells, fixed
(paraformaldehyde)
73 x 79 μ m

- Excitation: 266 nm (PicoQuant), 20 MHz, < 100 ps pulses
- Quartz objective, 40x, NA 0.6, glycerine
- Optical filters: Z266RDC, longpass 300 nm
- Detection: Ultrabialkali PMT (42 % @ 350 nm)

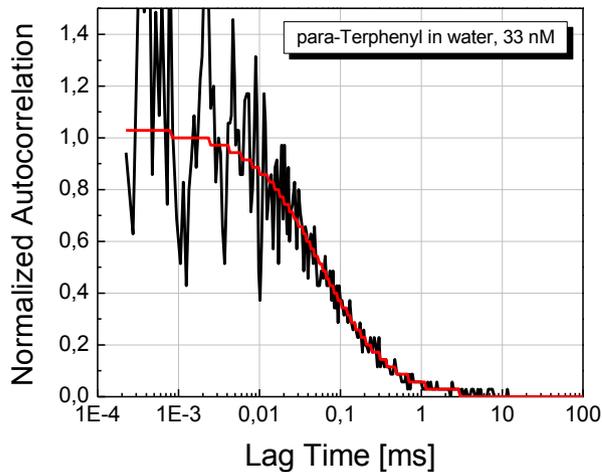
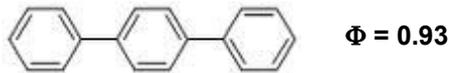
*Sample courtesy of Astrid Tannert,
University of Leipzig, Germany*

UV MicroTime 200: FCS

Benchmark: FCS in the deep UV *No commercially supported application*

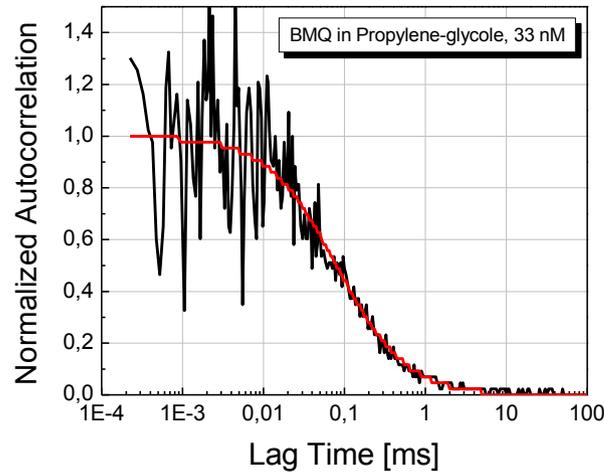
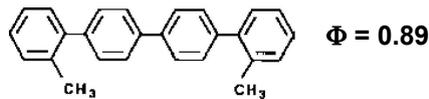
Dyes with high quantum efficiency in the UV: laser dyes as single emitters

para-Terphenyl



Diffusion time: $\tau(\text{diff}) = 57 \mu\text{s}$
Molecular brightness: 510 Hz per molecule
Ca. 220 μW , 10 min data acquisition

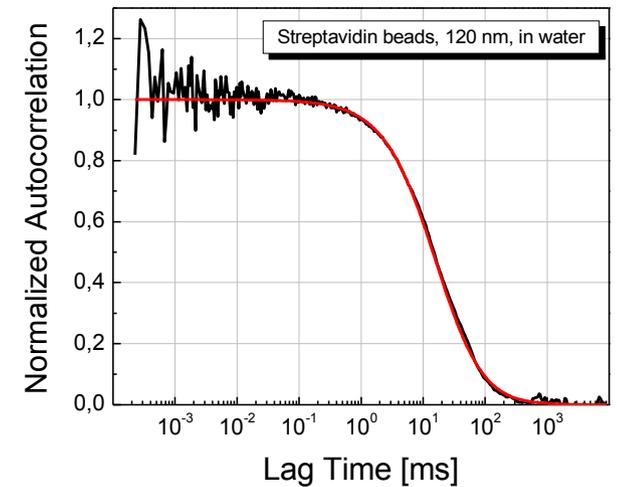
BMQ (2,2'-Dimethyl-p-terphenyl)



Diffusion time: $\tau(\text{diff}) = 79 \mu\text{s}$
Molecular brightness: 460 Hz per molecule
Ca. 280 μW , 15 min data acquisition

Streptavidin-coated beads (\varnothing 120 nm)

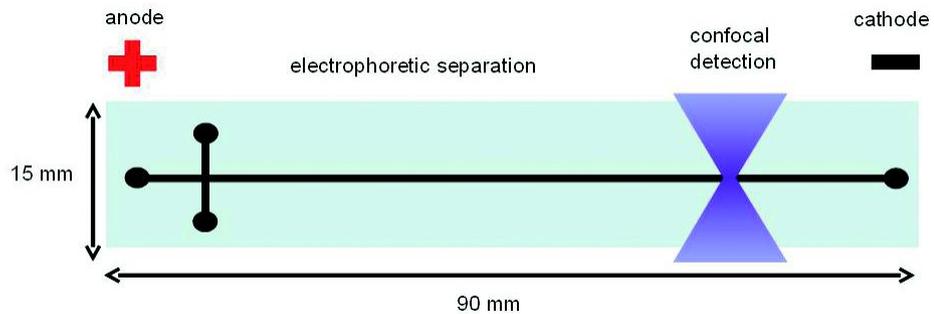
Approx. 1000 streptavidin per bead
(1 streptavidin = 24 tryptophans)



Diffusion time: $\tau(\text{diff}) = 16 \text{ ms}$
Molecular brightness: 1.2 kHz per bead
Ca. 200 μW , 5 min data acquisition

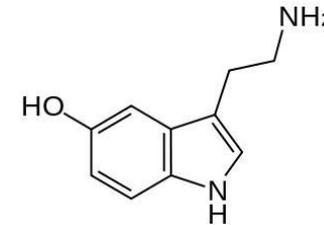
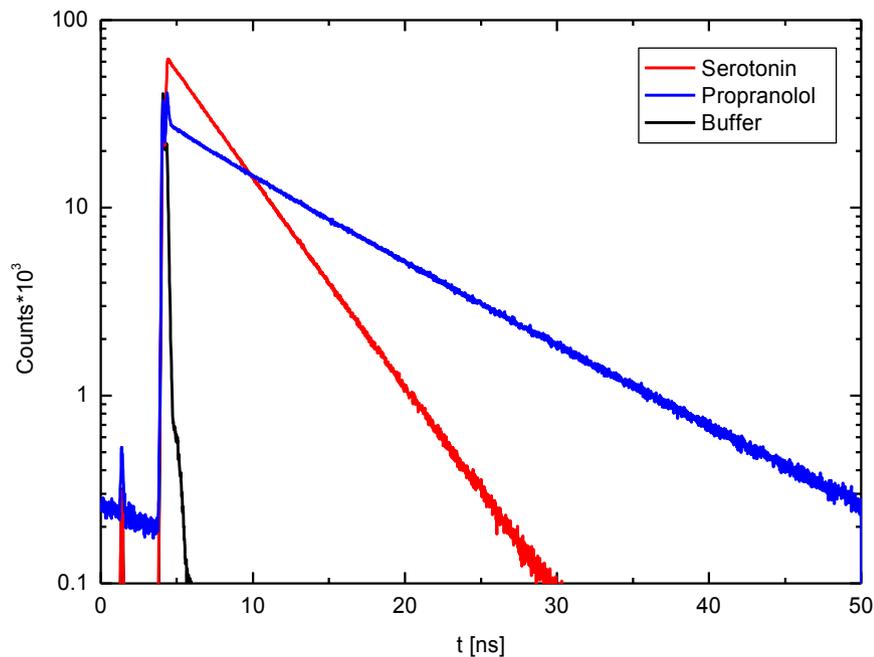
- Excitation: 266 nm (PicoQuant), 20 MHz, < 100 ps pulses
- Quartz objective, 40x, NA 0.6, glycerine
- Optical filters: Z266RDC, longpass 300 nm
- Detection: Ultrabialkali PMT (42 % @ 350 nm)

Microchip Electrophoretic Separation and Label Free Detection of Aromatic Analytes

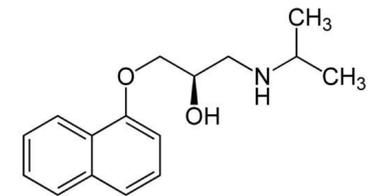


Substances in the microchannel (20 x 50 micron) are separated in the electric field and detected with a confocal microscope set-up.

Fluorescence decay curves are gathered on-the-fly and average lifetimes can be determined for each substance separately.



Serotonin



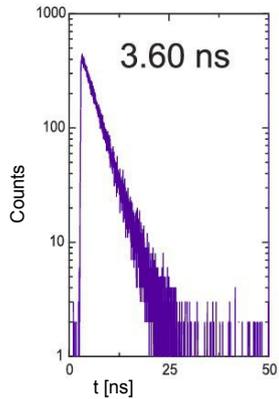
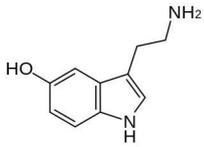
Propranolol

Time-resolved fluorescence decays of **serotonin** and **propranolol** (concentrations 250 μ M in buffer, Integration time 60 s).

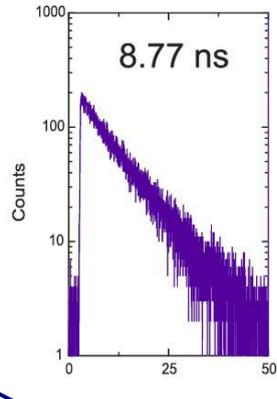
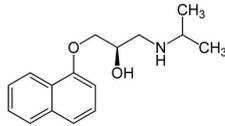
Max. 5 % deviation in lifetime determinations when compared to measurements in a mixture.

Electrophoretic Separation of Small Aromatics

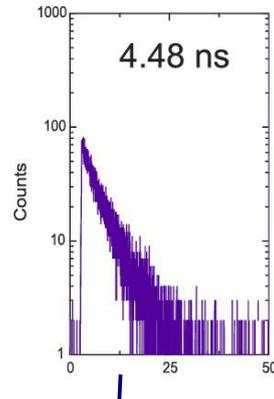
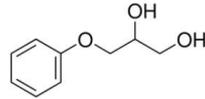
Serotonin



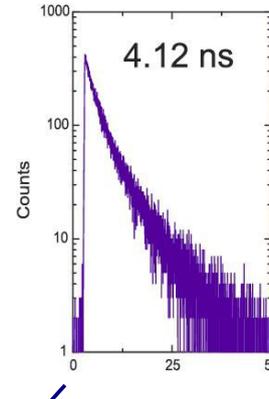
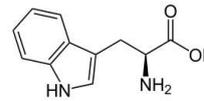
Propranolol



3-Phenoxy-1,2-Propanediol

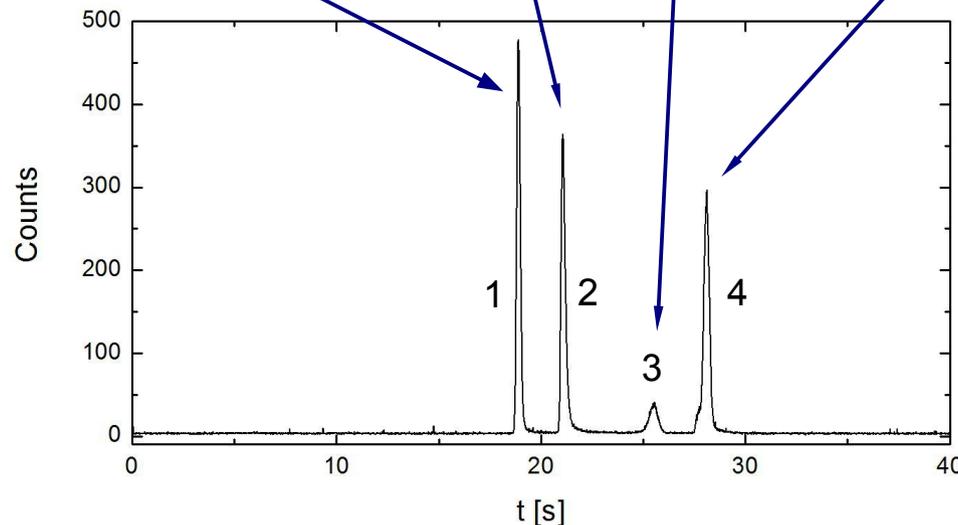


Tryptophan



- Excitation: 266 nm (Cougar, Time Bandwidth), 20 MHz, 10 ps pulses
- Quartz Objective, 40x, NA 0.8
- Optical filters: z266RDC, bandpass 350/50, DUG11x (285-365 nm)
- Detector: super-bialkali PMT

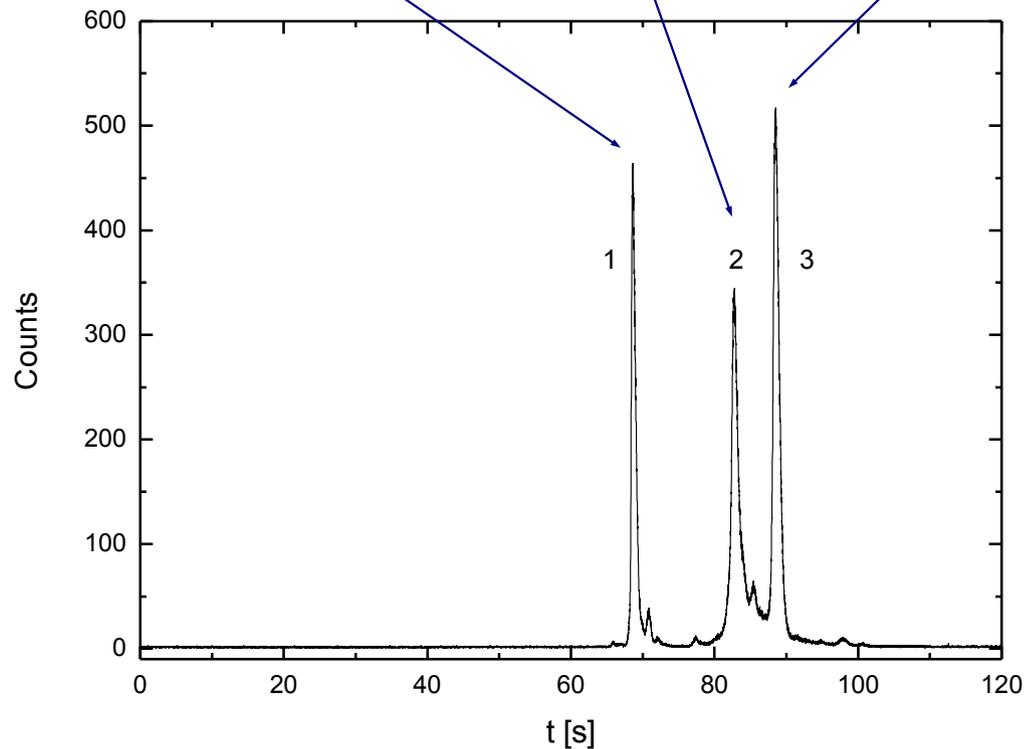
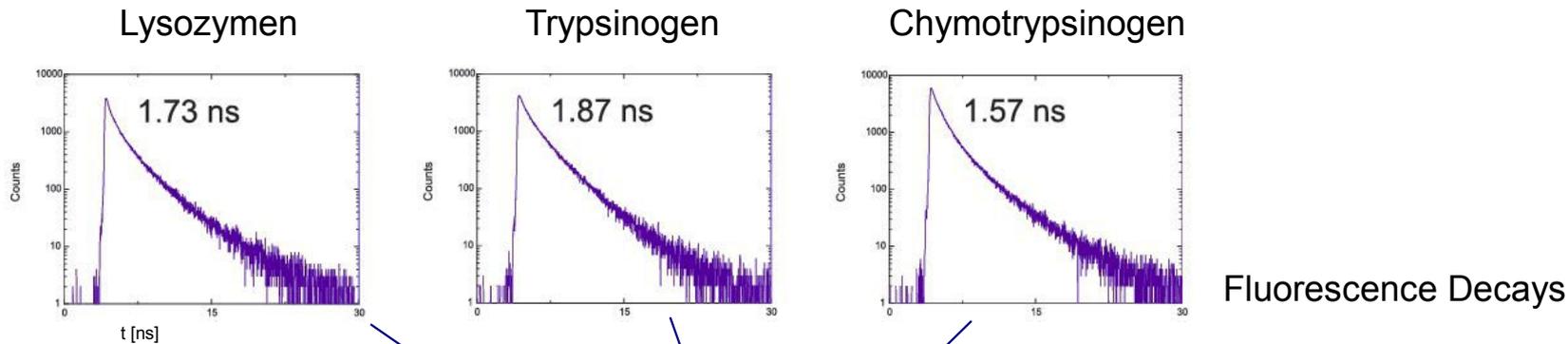
Fluorescence decays



Electropherogram:

188 μM serotonin (1), 135 μM propranolol (2), 238 μM 3-phenoxy-1,2-propanediol (3) and 198 μM tryptophan (4).
Separation Buffer: 5 mM borate, pH 9.2, effective separation length 5.0 cm

Electrophoretic Separation of Proteins



Electropherogram

23.3. μM lysozyme (1), 13.9 μM trypsinogen (2) and 13.0 μM chymotrypsinogen (3).
Separation buffer: 40 mM phosphate, pH 3.0
Microfluidic Channels were coated with 0.01 % w/v hydroxypropylmethylcellulose.
Effective separation length: 4.5 cm

- Excitation: 266 nm (Cougar, Time Bandwidth) 20 MHz, 10 ps pulses, 5nJ
- Quartz Objective, 40x, NA 0.8
- Optical filters: z266RDC, bandpass 350/50, DUG11x (285-365 nm)
- Detector: super-bialkali PMT

R. Beyreiss et al., Electrophoresis 2011, 32, 3108-3114

Further Informations

See specifications on our website or in the brochure.

Check our website for [training courses](#) on FLIM, FCS and Time-Correlated Single Photon Counting!

Share your experiences with the community in the PicoQuant forum at: <http://forum.picoquant.com/>

For further info on:

- Applications*
- Possible configurations*
- Pricing information*

please contact PicoQuant at info@picoquant.com